MGH Vector Core

Viral infection Protocol

Day 1: Plate 250,000-500,000 cells in one well of a 6-well plate.

Day 2: Mix 100 µL un-conentrated virus (or 2 µL concentrated virus) in a tube with 900 µL regular growth media and 1 µL polybrene (10 mg/mL stock). Vortex. Aspirate media from your cells. Replace with virus-containing media.

Day 3: If your construct contains a fluorescent reporter, check cells. Please note that it can take up to two days for cells to transduce. Depending on the confluency of your cells, remove virus-containing media, and either refresh with new media or passage your cells.

Viral Spinfection Protocol

Day 1: Plate 250,000-500,000 cells in one well of a 6-well plate.

Day 2: Mix 100 µL un-conentrated virus (or 2 µL concentrated virus) in a tube with 1.4 mL regular growth media and 1 µL polybrene (10 mg/mL stock). Vortex. Aspirate media from your cells. Replace with virus-containing media. Spin plate at 500*g* for 90 minutes at 32°C. Incubate cells overnight.

Day 3: If your construct contains a fluorescent reporter, check cells. Please note that it can take up to two days for cells to transduce. Depending on the confluency of your cells, remove virus-containing media, and either refresh with new media or passage your cells.

Note: Most experiments call for a mulplicity of infection (MOI) of 10, at which virtually 100% of cells in culture will be infected. We recommend adding 100 µL of your virus to your cells. However, you may use the following formula to determine your MOI with more precision.

MOI = (volume of virus in mL) x (viral titer) / (cell number)

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